

Inflammatory bowel disease: is it a primary immunodeficiency?

Erik Glocker · Bodo Grimbacher

Received: 13 September 2011 / Revised: 13 September 2011 / Accepted: 13 September 2011 / Published online: 14 October 2011
© Springer Basel AG 2011

Abstract Inflammatory bowel diseases (IBD) such as ulcerative colitis and Crohn's disease are chronic and relapsing conditions, characterized by abdominal pain, diarrhea, bleeding and malabsorption. IBD has been considered a hyperinflammatory state due to disturbed interactions between the immune system and the commensal bacterial flora of the gut. However, there is evidence that Crohn's disease might be the consequence of a reduced release of pro-inflammatory cytokines and an impaired acute inflammatory response, thereby suggesting that IBD might be an immunodeficiency rather than an excessive inflammatory reaction. This theory has been supported by observations in patients with primary immunodeficiencies such as the Wiskott–Aldrich syndrome and IPEX (immunodysregulation, polyendocrinopathy, enteropathy, X-linked syndrome). In contrary, defects in the anti-inflammatory down-regulation of the immune response as they are seen in patients with Mendelian defects in the IL10 signaling pathway support the hyper-inflammatory theory. In this review, we describe and

discuss primary immunodeficiencies associated with IBD and show that the bowel is a highly sensitive indicator of dysregulations, making IBD a model disease to study and identify key regulators required to balance the human mucosal immune system.

Keywords Inflammatory bowel disease · Immunodeficiency · IL10 deficiency · IL10R deficiency · Crohn's disease · Wiskott–Aldrich syndrome · CGD · IPEX

Introduction

Inflammatory bowel disease (IBD) is chronic in nature with a relapsing course and is accompanied by abdominal pain, diarrhea, bleeding, and malabsorption [1–3]. It comprises Crohn's disease (CD) and ulcerative colitis (UC) as well as indeterminate colitis with overlapping features of CD and UC [3]. IBD usually manifests in the second or third decade of life [4] but may also present in infancy, often with a severe and therapy resistant course of the disease [3]. IBD affects about 1.4 million people in the USA and 2.2 million in Europe [5, 6].

In general, IBD is suggested to result from disturbed interactions between the immune system and commensal bacteria of the gut [4, 6–8]. This theory is substantially backed by murine models showing that colitis does not develop in gnotobiotic mice, but emerges on reconstitution of the gut flora [4, 9, 10].

The intestinal integrity is maintained by several factors including the epithelial cell layer, mucus-secreting goblet cells, antimicrobial peptides-producing Paneth cells, IgA-releasing plasma cells, and gut-associated lymphoid tissue such as Peyer's patches [11, 12]. The chronic stimulation by

E. Glocker (✉)
Institute of Medical Microbiology and Hygiene,
University Medical Centre Freiburg, Hermann-Herder-Str. 11,
79104 Freiburg, Germany
e-mail: erik-oliver.glocker@uniklinik-freiburg.de

B. Grimbacher
Centre of Chronic Immunodeficiency, University Medical
Centre Freiburg, Breisacher Straße 177 - 2nd floor,
79106 Freiburg, Germany

B. Grimbacher (✉)
Department of Immunology, University College London
Medical School (Royal Free Campus),
Rowland Hill Street, London NW3 2PF, UK
e-mail: b.grimbacher@ucl.ac.uk

Table 1 Select number of genes associated with increased susceptibility to IBD (after van Limbergen et al. [18])

Function	Genes
Autophagy	<i>ATG16LI</i> , <i>IRGM</i> , <i>LRRK2</i>
Pattern-recognition receptors	<i>CARD9</i> , <i>NOD2</i> , <i>TLR4</i>
Th17 cell differentiation	<i>CCR6</i> , <i>ICOSLG</i> , <i>IL23R</i> , <i>JAK2</i> , <i>STAT3</i>
Maintenance of the epithelial barrier	<i>DLG5</i> , <i>DMBT1</i> , <i>ITLN1</i> , <i>OCTN1&2</i> , <i>ORMDL3</i> , <i>PTGER4</i> , <i>XPB1</i>
Shaping immune responses	<i>HLA region</i> , <i>IL12B</i> , <i>IL18RAP</i> , <i>IRF5</i> , <i>MST1</i> , <i>NKX2-3</i> , <i>PTPN22</i> , <i>TNFSF15</i>

ATG16LI: autophagy-related 16-like protein; *IRGM*: immunity-related GTPase family M; *LRRK2*: leucine-rich repeat kinase 2; *CARD9*: Caspase-recruitment domain containing protein 9; *NOD2*: nucleotide-binding oligomerization domain protein 2; *TLR4*: Toll-like receptor 4; *CCR6*: chemokine (C–C motif) receptor 6; *ICOSLG*: inducible T-cell co-stimulator ligand; *IL23R*: interleukin 23 receptor; *JAK2*: Janus kinase 2; *STAT3*: signal transducer and activator of transcription 3; *DLG5*: *Drosophila* discs large homologue; *DMBT1*: deleted in malignant brain tumors 1; *ITLN1*: intelectin 1; *OCTN1&2*: organic cation transporter 1&2; *ORMDL3*: ORM1-like 3; *PTGER4*: prostaglandin receptor EP4; *XPB1*: X-Box binding protein 1; *IL12B*: interleukin 12- β ; *IL18RAP*: IL18 receptor accessory protein; *IRF5*: interferon regulatory factor 5; *MST1*: macrophage stimulating protein 1; *NKX2-3*: NK2 transcription factor related, locus 3; *PTPN22*: protein tyrosine phosphatase, nonreceptor type 22; *TNFSF15*: tumor necrosis factor superfamily member 15

the resident intestinal flora and food antigens requires tight control mechanisms and involves IL-10 producing regulatory T (Treg) cells, which are supposed to be key players in maintaining the immune balance [13]. Mice lacking Treg cells suffer from fatal multiorgan inflammation; and IL-10 deficient mice die of wasting disease and colitis [14–16].

The genetic background of IBD has been in focus for several years, and genome-wide genetic linkage and association studies have underscored the genetic complexity of IBD and identified a wealth of genes that may render individuals more susceptible to IBD [17, 18]. Polymorphisms or mutations in genes involved in autophagy such as *ATG16LI* (autophagy-related 16-like protein), *IRGM* (immunity-related GTPase family M); [19–22] intra- and extracellular pattern recognition receptors such as *NOD2* (nucleotide-binding oligomerization domain protein 2) or *TLR4* (Toll-like receptor 4); [23–26] Th17 cell differentiation such as *IL23R* (interleukin 23 receptor) and *STAT3* (signal transducer and activator of transcription 3); [19, 27–30] maintenance of the intestinal epithelium such as *DLG5* (*Drosophila* discs large homologue), *OCTN1&2* (organic cation transporter) and *XPB1* (X-Box-binding protein 1); [31, 32] and shaping immune responses such as *IL12B* (interleukin 12- β) and *IL18RAP* (IL18 receptor accessory protein) [19, 33] have been shown to be associated with or increase susceptibility to IBD (summarized in Table 1).

In contrast to the hypothesis, that IBD is a complex multigenic disorder, evidence arose that certain forms of IBD may (1) be congenital/inborn Mendelian traits as demonstrated in IL-10- and IL-10 receptor deficiency; and (2) reflect an immunodeficient state of the host as shown in primary immunodeficiencies (PID) such as Wiskott–Aldrich syndrome or chronic granulomatous disease (CGD) [34–38]. In this review, we will discuss the cases when a PID should be suspected in a patient with IBD.

Crohn's disease

CD is characterized by a segmental transmural inflammation of the gut with the formation of non-caseating granuloma, which consist of macrophages, epithelioid, and giant cells. CD may affect any site of the intestine, and involvement of the terminal ileum is most common [1, 2, 4].

The prevailing hypothesis on the pathogenesis suggests that CD, like other forms of IBD, may be the consequence of a hyperinflammatory state due to an inappropriate immune response to the intestinal flora [4, 40].

In the 1970s, the first evidence emerged that CD may be the consequence of an impaired acute inflammatory response with an incomplete removal of bacteria and foreign material in the gut [41]. This concept of CD as an immunodeficiency was backed and extended by a study of Marks et al., who showed that Crohn's patients had a weaker efflux of neutrophils into skin windows as compared to healthy controls or patients suffering from UC or rheumatoid arthritis [42]. Patients also revealed reduced amounts of pro-inflammatory cytokines such as IL-1 β or IL-8 and a massively impaired migration of neutrophils in lesions of the rectum or ileum upon taking tissue samples [42]. In contrast to healthy individuals, CD's patients revealed an abnormal low change in blood flow on subcutaneous injection with heat-killed *Escherichia coli*, confirming a defective inflammatory response.

Recent work by Smith et al. [43] showed that the migration of ^{111}In labeled neutrophils and the clearance of subcutaneously injected ^{32}P labeled *E. coli* were substantially impaired in CD patients, the blood flow and the removal of bacteria was dose-dependent. Macrophages from CD patients had a substantially impaired release of pro-inflammatory cytokines such as TNF- α , IL-4, IL-5, IL-13, IL-15 and IFN- γ on stimulation with heat-killed *E. coli* or Toll-like receptor agonists when compared to healthy controls or patients suffering from UC. The authors' investigations revealed that intracellular levels of TNF- α in macrophages from CD patients were significantly lower than in healthy individuals, even though transcription and intracellular mRNA levels were normal. The addition of Brefeldin-A, an inhibitor of the transport of proteins

from the endoplasmic reticulum to the Golgi-apparatus, prevented the degradation of TNF- α , indicating that the decreased TNF- α release may be due to damaging events in the lysosomal compartments [43].

The lack of an initial acute inflammation with the reduced elimination of foreign material and bacteria may result in a chronic inflammation with T cell-mediated granuloma formation, which are supposed to confine hazardous material and protect the intestine from bacterial spreading [43, 44]. The development of a CD-like enterocolitis in CGD supports the theory that CD is an immunodeficiency due to impaired pro-inflammatory responses. The continuous secretion of pro-inflammatory cytokines by cells like macrophages that form the granuloma, sustains the chronic inflammation and finally results in extensive tissue damage. A general immunodeficiency of macrophages may also be an explanation for the occurrence of other CD manifestations such as arthritis or eye- and skin lesions [43–45].

IL-10- and IL-10 receptor deficiency

IL-10 is secreted by several cell types including monocytes, macrophages and dendritic cells, T cells, B cells, granulocytes, epithelial cells, keratinocytes, and mast cells [46]. It is critical in maintaining the balance of the immune system, restricts and terminates immune responses by limiting the secretion of pro-inflammatory cytokines such as TNF- α , IL-1, IL-6 and IL-12, and controls both the differentiation and proliferation of macrophages, T and B cells [46–49]. The relevance of IL-10 and its signaling for maintaining intestinal immune homeostasis became evident in murine models: Both IL-10- (*Il10*^{−/−}) and IL-10-receptor (*Il10rb*^{−/−})-deficient mice develop severe enterocolitis when the gastrointestinal flora develops [7, 50, 51].

Due to its unique role in balancing the immune system, IL-10 has always been in the focus of IBD research. Genetic sequence variants in IL-10 were shown to contribute to susceptibility to ulcerative colitis [52], and mutations in the IL-10 leader sequence were found to modify the IL-10 release in patients with CD [53]. A study by Noguchi et al. [54] demonstrated that a mutant NOD2 protein that has been associated with CD inhibits the ribonucleoprotein hnRNP-A1 and thus actively suppress the transcription of IL-10. NOD2/CARD15 is an intracellular sensor that recognizes bacterial peptidoglycan and has been shown to be a risk factor for the development of IBD [23, 55, 56].

By analyzing two families with an autosomal-recessive inherited-type of enterocolitis, we recently demonstrated that IBD may be a monogenic disorder [34]. The patients presented within the first year of life with enterocolitis and

perianal disease, and the formation of multiple abscesses and enterocutaneous fistula, which required several surgical interventions. Histopathology revealed ulcerations of the intestinal mucosa with inflammatory infiltrates of the epithelium and the formation of abscesses extending to the *muscularis propria*. Additionally, the patients suffered from chronic folliculitis and recurrent respiratory infections. The patients were treated with various anti-inflammatory drugs, including steroids, methotrexate, thalidomide, and anti-TNF- α monoclonal antibodies, but none of these therapies induced remission or long-term improvement [34].

Our genetic studies identified two patients with mutations in *IL10RA* (encoding the α -unit of the IL-10 receptor, IL10R1), which resulted in amino acid exchanges at position 84 (Thr84Ile) or 141 (Gly141Arg). The other patients, two siblings, harbored a mutation affecting the *IL10RB* gene, [encoding the β -unit (IL10R2)], which resulted in a premature stop codon (Try159Stop) [34]. Apart from the four patients published [34], further patients with mutations in the IL-10 receptor have been found: two additional patients were identified in the UK, four in Germany, one in the US, one in Canada and one in France (privileged communication; unpublished data).

The IL-10 receptor is a tetramer consisting of two α -chains (IL10R1) and two β -chains (IL10R2) molecules [46, 57]. Upon binding of IL-10 dimers to the α -chains, the accessory β -chain are recruited and a functional IL10 receptor tetramer assembled. This gives rise to the activation of Janus tyrosine kinases (JAK)1 and Tyk2, rapid phosphorylation of STAT3, and induction of STAT3-dependent genes such as the suppressor of cytokine signaling (SOCS)-3 gene [57–60].

The mutations found in these patients abolished IL-10-induced STAT3 phosphorylation and prevented IL-10-mediated inhibition of TNF- α release in LPS-stimulated macrophages and peripheral blood mononuclear cells (PBMCs) [34].

In contrast to IL10R1, which is unique to the IL-10 receptor, IL10R2 is also a component of the receptors for IL-22, IL-26, IL-28A, IL-28B, and IL-29 and expressed on a wide range of non-immune cells such as epithelial cells and keratinocytes [61–63]. In particular, IL-22 may add to the severe phenotype in IL10R2-deficient patients, since it was shown to protect against colitis and improved colitis in mice [64, 65]. IL-22 upregulates expression of the antimicrobial proteins RegIII β and γ and enhances mucus production in murine colonic epithelial cells, thereby maintaining the epithelial barrier and protecting from infections with intestinal bacterial pathogens [64, 66].

The recurrent and frequent folliculitis in the IL10R2-deficient patients may be at least in part attributed to a lack of IL-22 signaling, which is supposed to control immunity

of the skin by up-regulating the expression of the β -defensins 2 and 3 and the antimicrobial heterodimer S100A8/9 in keratinocytes [67–69]. The λ -interferons IL-28A, IL-28B, and IL-29 are known primarily to confer antiviral protection [70, 71], but since other vital antiviral defenses such as the IFN type I and II signaling pathways are still intact, this shortcoming may be of minor relevance.

Due to the dramatic clinical situation, a hematopoietic stem cell transplantation (HSCT) was performed in one of the patients with an IL10R2 mutation, which proved to be successful and cured the patient [34]. The sustained success of the HSCT suggests that IL-10 signaling in hematopoietic cells rather than signaling via IL-22/IL-26/IFN- λ in non-haematopoietic cells was critical to induce remission.

Most recently, we described two other patients with Crohn's-like disease and the formation of perianal and rectovaginal fistulae [35]. Endoscopy and histopathology revealed extensive ulceration of the ileum and focal active colitis with neutrophils infiltrating the surface epithelium. Both patients carried homozygous loss-of-function mutations in *IL-10* itself, leading to an amino acid exchange at codon 113 (Gly113Arg), which most likely impeded dimerization of IL-10 as shown by molecule modeling. The mutated IL-10 failed to induce STAT3 phosphorylation and to inhibit LPS-mediated TNF- α release in PBMCs [35].

The dramatic clinical findings in patients with either IL-10 or IL-10 receptor mutations emphasize the importance of this cytokine in controlling the intestinal immune system and show that the loss of IL-10 signaling cannot be compensated by any other pathway.

Immunodysregulation, polyendocrinopathy, enteropathy, X-linked syndrome (IPEX)

Due to the massive amount of resident colonizing bacteria in the gut, highly sophisticated mechanisms are required to maintain the integrity of the intestine, to balance the intestinal immune system and to sense and distinguish the physiological flora from invading pathogens. A plethora of different cells contribute to this challenge including epithelial cells, Paneth cells, goblet cells, and cells of the adaptive immune system [11].

A key player in maintaining the intestinal immune homeostasis are CD4⁺/CD25⁺/FoxP3⁺ Treg cells and FoxP3-Tr1-like cells, which both are important sources of IL-10 and suppress pro-inflammatory action [11, 72, 73]. Several murine models show the need for Treg cells: FoxP3⁺-deficient mice develop severe multiorgan inflammation, which improves upon transfer of FoxP3⁺ positive Treg cells [14], and Treg cells have been shown not only to control intestinal inflammation but also to cure manifest colitis [73, 74]. The majority of CD4⁺/CD25⁺/FoxP3⁺

Treg cells develop in the thymus, where their T cell receptors bind self-peptides with moderate affinity, which results in activation of NF- κ B [11, 75, 76]. Co-stimulation via CD28 and IL-2 or IL-15 activate STAT5 and induce expression of FoxP3 [77]. Some Treg cells then may migrate from the thymus to the gastrointestinal tract and recognize intestinal antigens, thereby preventing excessive immune reactions [11, 78].

Patients with mutation in the *FOXP3* gene, located on the X-chromosome, suffer from IPEX syndrome that is characterized by a lack of Treg cells, leading to autoimmune lymphoproliferation and multiple autoimmune disorders [79–82]. If not fatal in early childhood, IPEX patients usually present with insulin-dependent diabetes mellitus, failure to thrive, skin disease such as eczema, hypo- or hyperthyroidism and recurrent infections including meningitis, pneumonia, and septicemia [82–85]. The predominant clinical feature is an autoimmune enteropathy that may mimic CD, UC, or celiac disease [83, 86]. The small intestine reveals complete or partial villous atrophy, the large intestine is frequently involved and shows inflammatory infiltrates with CD3⁺ cells and plasma cells [83, 86, 87]. The frequently atrophic thymus may show a substantial reduction of Hassall's corpuscles, which secrete thymic stromal lymphopoietin and activate dendritic cells, which then induce differentiation of Treg cells [86, 88, 89].

Chronic granulomatous disease (CGD)

Several congenital disorders affecting the function phagocytes including glycogen storage disease-1b, Chediak-Higashi- and Hermansky-Pudlak-syndrome, cyclical neutropenia, and leukocyte adhesion deficiency 1 have been shown to be associated with forms of enterocolitis [39, 90]. A prototype of neutrophil disorder that resembles CD is chronic granulomatous disease (CGD). CGD is a rare disorder in which the respiratory burst of neutrophils is dramatically impaired due to mutations in the components gp91^{Phox}, p47^{Phox}, p67^{Phox} and p22^{Phox} of the NADPH oxidase complex [91]. Together with p22rac, these proteins form a complex in the wall of the phagocytic vacuole [39, 92], which enables the electron transport from NADPH on O₂ to generate microbicidal O₂⁻ and H₂O₂ in the vacuole, which are required for intracellular killing of microbes or digestion of other material [39, 93]. The most frequent form of CGD is X-linked with mutations in gp91^{Phox}, accounting for about 65% of cases [39]. The impaired killing of microorganisms including possible pathogens render patients with CGD highly susceptible to severe recurrent bacterial and fungal infections, in particular *Staphylococcus aureus*, *Salmonella* spp., and *Aspergillus* spp [94, 95].

A hallmark of CGD is the formation of abnormal inflammatory granuloma that may result in colitis and may obstruct organ systems such as the urinary tract [38, 95–97]. The formation of granuloma in the absence of infectious agents may be the result of an incapability of neutrophils to digest foreign material. This is followed by a second chronic stage of inflammation with the formation of granuloma, which results in inflammatory bowel disease that resembles CD [38, 39].

Wiskott–Aldrich syndrome

The Wiskott–Aldrich syndrome (WAS) is a rare immunodeficiency caused by mutations in the Wiskott–Aldrich syndrome protein (WASP) gene that is located on the X chromosome. WASP is exclusively expressed in hematopoietic cells and transduces signals from the cell surface to the actin cytoskeleton. The actin cytoskeleton is required to control cell–cell interaction, cell movement, cell signaling and cell division [98]. WASP is inactive in the cytoplasm due to an auto-inhibitory mechanism mediated by interaction between the verprolin–cofilin homology domains acidic region (VCA) and the GTPase-binding domain (GBD) [99]. Upon activation by the Rho GTPase cell division cycle 42 (Cdc42) or non-catalytic region of tyrosine kinase 1 (Nck1), this inhibition is abolished, enabling the VCA domain to bind the actin-related protein (ARP) 2/3 complex and stimulates actin polymerization [99–101]. Numerous mutations have been described to date, and there is a clear-cut correlation between the type of mutation and the clinical phenotype: if WASP is not expressed, patients present with the classic WAS, expression of mutated WASP results in X-linked thrombocytopenia, and mutation in the binding site of the small rho GTPase Cdc42 gives rise to X-linked neutropenia [102–105]. Recurrent infections due to immunodeficiency, eczema, microthrombocytopenia, and increased frequencies of autoimmune diseases are the clinical hallmarks of the Wiskott–Aldrich syndrome [102, 106, 107]. Autoimmune conditions are found in up to 40% of the patients and include hemolytic anemia, vasculitis, Henoch–Schönlein-like purpura and inflammatory bowel disease [102, 108], which may resemble UC and has been documented for up to 10% of WAS patients [109, 110]. WASP-deficient Treg cells showed impaired suppressive activity in vitro and failed to control autoimmunity in several murine models [99]. Why the other 90% of WAS patients do not suffer from IBD is unclear.

Conclusions

IBD comprises a group of diseases that may be caused by a variety of different irregularities of the immune system.

The occurrence of an IBD phenotype in WAS, IPEX, Artemis deficiency [111] and in particular CGD indicates that IBD may be the final stage of several complex immunodeficiencies. Recent work confirmed that a classical form of IBD, namely CD, may be an immunodeficiency due to a malfunction of macrophages with faulty release of cytokines and subsequent impaired acute inflammatory response to bacteria and foreign material in the intestine. This contradicts the previous prevailing hypothesis that CD is a hyper-inflammatory disorder due to an inadequate reaction of the innate and adaptive immune system to the flora of the gut and may suggest new therapeutic strategies.

The detection of mutations in the IL-10 signaling pathway demonstrates the importance of immunoregulatory factors that keep the intestinal immune system in balance; it also shows that in a subgroup of patients IBD may be monogenic and distinct from more complex forms like UC and CD. Even though the progress in IBD research has broadened our knowledge of the intestinal mucosal immunity and the components that keep it in balance, there are still many other factors that need to be identified to make us understand the complex eco-system of the gut.

References

- Podolsky DK (2002) Inflammatory bowel disease. *N Engl J Med* 347:417–429
- Baumgart DC, Carding SR (2007) Inflammatory bowel disease: cause and immunobiology. *Lancet* 369:1627–1640
- Romano C, Famiani A, Gallizzi R et al (2008) Indeterminate colitis: a distinctive clinical pattern of inflammatory bowel disease in children. *Pediatrics* 122:e1278–e1281
- Xavier RJ, Podolsky DK (2007) Unravelling the pathogenesis of inflammatory bowel disease. *Nature* 448:427–434
- Carter MJ, Lobo AJ (2004) Travis SP; IBD Section, British Society of Gastroenterology. Guidelines for the management of inflammatory bowel disease in adults. *Gut* 53(Suppl 5):V1–V16
- Engel MA, Neurath MF (2010) New pathophysiological insights and modern treatment of IBD. *J Gastroenterol* 45:571–583
- Spencer SD, Di Marco F, Hooley J et al (1998) The orphan receptor CRF2–4 is an essential subunit of the interleukin 10 receptor. *J Exp Med* 187:571–578
- Mizoguchi A, Mizoguchi E (2008) Inflammatory bowel disease, past, present and future: lessons from animal models. *J Gastroenterol* 43:1–17
- Elson CO, Cong Y, McCracken VJ, Dimmitt RA, Lorenz RG, Weaver CT (2005) Experimental models of inflammatory bowel disease reveal innate, adaptive, and regulatory mechanisms of host dialogue with the microbiota. *Immunol Rev* 206:260–276
- Onderdonk AB, Hermos JA, Bartlett JG (1977) The role of the intestinal microflora in experimental colitis. *Am J Clin Nutr* 30:1819–1825
- Barnes M, Powrie F (2009) Regulatory T cells reinforce intestinal homeostasis. *Immunity* 31:401–411
- Artis D (2008) Epithelial-cell recognition of commensal bacteria and maintenance of immune homeostasis in the gut. *Nat Rev Immunol* 8:411–420

13. Boden EK, Snapper SB (2008) Regulatory T cells in inflammatory bowel disease. *Curr Opin Gastroenterol* 24:733–741
14. Fontenot JD, Gavin MA, Rudensky AY (2003) Foxp3 programs the development and function of CD4⁺ CD25⁺ regulatory T cells. *Nat Immunol* 4:330–336
15. Li MO, Wan YY, Flavell RA (2007) T cell-produced transforming growth factor-beta1 controls T cell tolerance and regulates Th1- and Th17-cell differentiation. *Immunity* 26:579–591
16. Rubtsov YP, Rasmussen JP, Chi EY, Fontenot J, Castelli L, Ye X, Treuting P, Siewe L, Roers A, Henderson WR Jr, Muller W, Rudensky AY (2008) Regulatory T cell-derived interleukin-10 limits inflammation at environmental interfaces. *Immunity* 28:546–558
17. Cho JH (2008) The genetics and immunopathogenesis of inflammatory bowel disease. *Nat Rev Immunol* 8:458–466
18. Van Limbergen J, Wilson DC, Satsangi J (2009) The genetics of Crohn's disease. *Annu Rev Genomics Hum Genet* 10:89–116
19. Barrett JC, Hansoul S, Nicolae DL et al (2008) Genome-wide association defines more than 30 distinct susceptibility loci for Crohn's disease. *Nat Genet* 40:955–962
20. Hampe J, Franke A, Rosenstiel P et al (2007) A genome-wide association scan of nonsynonymous SNPs identifies a susceptibility variant for Crohn disease in ATG16L1. *Nat Genet* 39:207–211
21. Parkes M, Barrett JC, Prescott NJ et al (2007) Sequence variants in the autophagy gene *IRGM* and multiple other replicating loci contribute to Crohn's disease susceptibility. *Nat Genet* 39:830–832
22. Cadwell K, Liu JY, Brown SL et al (2008) A key role for autophagy and the autophagy gene *Atg16l1* in mouse and human intestinal Paneth cells. *Nature* 456:259–263
23. Hugot JP, Chamaillard M, Zouali H et al (2001) Association of NOD2 leucine-rich repeat variants with susceptibility to Crohn's disease. *Nature* 411:599–603
24. Economou M, Trikalinos TA, Loizou KT et al (2004) Differential effects of NOD2 variants on Crohn's disease risk and phenotype in diverse populations: a metaanalysis. *Am J Gastroenterol* 99:2393–2404
25. Pierik M, Joossens S, Van Steen K et al (2006) Toll-like receptor-1, -2, and -6 polymorphisms influence disease extension in inflammatory bowel diseases. *Inflamm Bowel Dis* 12:1–8
26. van Heel DA, Fisher SA, Kirby A et al (2004) Inflammatory bowel disease susceptibility loci defined by genome scan meta-analysis of 1952 affected relative pairs. *Hum Mol Genet* 13:763–770
27. Duerr RH, Taylor KD, Brant SR et al (2006) A genome-wide association study identifies IL23R as an inflammatory bowel disease gene. *Science* 314:1461–1463
28. Glas J, Seiderer J, Wetzke M et al (2007) rs1004819 is the main disease-associated IL23R variant in German Crohn's disease patients: combined analysis of IL23R, CARD15, and OCTN1/2 variants. *PLoS One* 2:e819
29. Fisher SA, Tremelling M, Anderson CA et al (2008) Genetic determinants of ulcerative colitis include the ECM1 locus and five loci implicated in Crohn's disease. *Nat Genet* 40:710–712
30. Taylor KD, Targan SR, Mei L et al (2008) IL23R haplotypes provide a large population attributable risk for Crohn's disease. *Inflamm Bowel Dis* 14:1185–1191
31. Stoll M, Corneliussen B, Costello CM et al (2004) Genetic variation in *DLG5* is associated with inflammatory bowel disease. *Nat Genet* 36:476–480
32. Kaser A, Lee AH, Franke A et al (2008) XBP1 links ER stress to intestinal inflammation and confers genetic risk for human inflammatory bowel disease. *Cell* 134:743–756
33. Zhernakova A, Festen EM, Franke L et al (2008) Genetic analysis of innate immunity in Crohn's disease and ulcerative colitis identifies two susceptibility loci harboring CARD9 and IL18RAP. *Am J Hum Genet* 82:1202–1210
34. Glocker EO, Kotlarz D, Boztug K et al (2009) Inflammatory bowel disease and mutations affecting the interleukin-10 receptor. *N Engl J Med* 361:2033–2045
35. Glocker EO, Frede N, Perro M, et al. Severe early-onset colitis due to a mutation in interleukin-10. *The Lancet* (accepted)
36. Akman IO, Ostrov BE, Neudorf S (1998) Autoimmune manifestations of the Wiskott–Aldrich syndrome. *Semin Arthritis Rheum* 27:218–225
37. Marks DJB, Segal AW (2008) Innate immunity in inflammatory bowel disease: a disease hypothesis. *J Pathol* 214:260–266
38. Marks DJB, Miyagi K, Rahman FZ et al (2009) Inflammatory bowel disease in CGD reproduces the clinicopathological features of Crohn's disease. *Am J Gastroenterol* 104:117–124
39. Rahman FZ, Marks DJB, Hayee BH et al (2008) Phagocytic dysfunction and inflammatory bowel disease. *Inflamm Bowel Dis* 14:1443–1452
40. Bouma G, Strober W (2003) The immunological and genetic basis of inflammatory bowel disease. *Nat Rev Immunol* 3:521–533
41. Segal AW, Loewi G (1976) Neutrophil dysfunction in Crohn's disease. *Lancet* 308:219–221
42. Marks DJB, Harbord MWN, MacAllister R et al (2006) Defective acute inflammation in Crohn's disease: a clinical investigation. *Lancet* 367:668–678
43. Smith AM, Rahman FZ, Hayee BH et al (2009) Disordered macrophage cytokine secretion underlies impaired acute inflammation and bacterial clearance in Crohn's disease. *J Exp Med* 206:1883–1897
44. Casanova JL, Abel L (2009) Revisiting Crohn's disease as a primary immunodeficiency of macrophages. *J Exp Med* 206:1839–1843
45. Ephgrave K (2007) Extra-intestinal manifestations of Crohn's disease. *Surg Clin North Am* 87:673–680
46. Moore KW, de Waal Malefyt R, Coffman RL, O'Garra A (2001) Interleukin-10 and the interleukin-10 receptor. *Annu Rev Immunol* 19:683–765
47. Bogdan C, Vodovotz Y, Nathan C (1991) Macrophage deactivation by interleukin 10. *J Exp Med* 174:1549–1555
48. de Waal Malefyt R, Haanen J, Spits H et al (1991) Interleukin-10 (IL-10) and viral IL-10 strongly reduce antigen-specific human T cell proliferation by diminishing the antigen-presenting capacity of monocytes via down regulation of class II major histocompatibility complex expression. *J Exp Med* 174:915–924
49. Williams LM, Ricchetti G, Sarma U et al (2004) Interleukin-10 suppression of myeloid cell activation—a continuing puzzle. *Immunology* 113:281–292
50. Kühn R, Lohler J, Rennick D et al (1993) Interleukin-10-deficient mice develop chronic enterocolitis. *Cell* 75:263–274
51. Berg DJ, Kühn R, Rajewsky K et al (1995) Interleukin-10 is a central regulator of the response to LPS in murine models of endotoxic shock and the Shwartzman reaction but not endotoxin tolerance. *J Clin Invest* 96:2339–2347
52. Franke A, Balschun T, Karlsen TH et al (2008) Sequence variants in *IL10*, *ARPC2* and multiple other loci contribute to ulcerative colitis susceptibility. *Nat Genet* 40:1319–1323
53. van der Linde K, Boor PP, Sandkuijl LA et al (2003) A Gly15Arg mutation in the interleukin-10 gene reduces secretion of interleukin-10 in Crohn disease. *Scand J Gastroenterol* 38:611–617
54. Noguchi E, Homma Y, Kang X et al (2009) A Crohn's disease-associated NOD2 mutation suppresses transcription of human

- IL10 by inhibiting activity of the nuclear ribonucleoprotein hnRNP-A1. *Nat Immunol* 10:471–479
55. Girardin SE, Hugot JP, Sansonetti PJ (2003) Lessons from Nod2 studies: towards a link between Crohn's disease and bacterial sensing. *Trends Immunol* 24:652–658
 56. Ogura Y, Bonen DK, Inohara N et al (2001) A frameshift mutation in NOD2 associated with susceptibility to Crohn's disease. *Nature* 411:603–606
 57. Donnelly RP, Dickensheets H, Finbloom DS (1999) The interleukin-10 signal transduction pathway and regulation of gene expression in mononuclear phagocytes. *J Interferon Cytokine Res* 19:563–573
 58. Kotenko SV, Krause CD, Izotova LS et al (1997) Identification and functional characterization of a second chain of the interleukin-10 receptor complex. *The EMBO J* 16:5894–5903
 59. O'Shea JJ, Murray PJ (2008) Cytokine signaling modules in inflammatory responses. *Immunity* 28:477–487
 60. Williams L, Bradley L, Smith A, Foxwell B (2004) Signal transducer and activator of transcription 3 is the dominant mediator of the anti-inflammatory effects of IL-10 in human macrophages. *J Immunol* 172:567–576
 61. Wolk K, Sabat R (2006) Interleukin-22: A novel T- and NK-cell derived cytokine that regulates the biology of tissue cells. *Cytokine & Growth Factor Rev* 17:367–380
 62. Commins S, Steinke JW, Borish L (2008) The extended IL-10 superfamily: IL-10, IL-19, IL-20, IL-22, IL-24, IL-26, IL-28, and IL-29. *J Allergy Clin Immunol* 121:1108–1111
 63. Donnelly RP, Sheikh F, Kotenko SV, Dickensheets H (2004) The expanded family of class II cytokines that share the IL-10 receptor-2(IL-10R2) chain. *J Leukoc Biol* 76:314–321
 64. Sugimoto K, Ogawa A, Mizoguchi E et al (2008) IL-22 ameliorates intestinal inflammation in a mouse model of ulcerative colitis. *J Clin Invest* 118:534–544
 65. Zenewicz LA, Yancopoulos GD, Valenzuela DM et al (2008) Innate and adaptive interleukin-22 protects mice from inflammatory bowel disease. *Immunity* 29:947–957
 66. Zheng Y, Valdez PA, Danilenko DM et al (2008) Interleukin-22 mediates early host defense against attaching and effacing bacterial pathogens. *Nat Med* 14:282–289
 67. Wolk K, Kunz S, Witte E, Friedrich M, Asadullah K, Sabat R (2004) IL-22 increases the innate immunity of tissues. *Immunity* 21:241–254
 68. Wolk K, Witte E, Wallace E, Döcke WD, Kunz S, Asadullah K, Volk HD, Sterry W, Sabat R (2006) IL-22 regulates the expression of genes responsible for antimicrobial defense, cellular differentiation, and mobility in keratinocytes: a potential role in psoriasis. *Eur J Immunol* 36:1309–1323
 69. Kolls JK, McCray PB Jr, Chan YR (2008) Cytokine-mediated regulation of antimicrobial proteins. *Nat Rev Immunol* 8:829–835
 70. Kotenko SV, Gallagher G, Baurin VV et al (2003) IFN- λ s mediate antiviral protection through a distinct class II cytokine receptor complex. *Nat Immunol* 4:69–77
 71. Sheppard P, Kindsvogel W, Xu W et al (2003) IL-28, IL-29 and their class II cytokine receptor IL-28R. *Nat Immunol* 4:63–68
 72. Maloy KJ, Salaun L, Cahill R et al (2003) CD4 + CD25 + T(R) cells suppress innate immune pathology through cytokine-dependent mechanisms. *J Exp Med* 197:111–119
 73. Read S, Malmström V, Powrie F (2000) Cytotoxic T lymphocyte-associated antigen 4 plays an essential role in the function of CD25(+)CD4(+) regulatory cells that control intestinal inflammation. *J Exp Med* 192:295–302
 74. Mottet C, Uhlig HH, Powrie F (2003) Cutting edge: cure of colitis by CD4+ CD25+ regulatory T cells. *J Immunol* 170:3939–3943
 75. Feuerer M, Hill JA, Mathis D, Benoist C (2009) Foxp3+ regulatory T cells: differentiation, specification, subphenotypes. *Nat Immunol* 10:689–695
 76. Josefowicz SZ, Rudensky A (2009) Control of regulatory T cell lineage commitment and maintenance. *Immunity* 30:616–625
 77. Lio CW, Hsieh CS (2008) A two-step process for thymic regulatory T cell development. *Immunity* 28:100–111
 78. Powrie F, Leach MW, Mauze S et al (1993) Phenotypically distinct subsets of CD4+ T cells induce or protect from chronic intestinal inflammation in C. B-17 scid mice. *Int Immunol* 5:1461–1471
 79. Bennett CL, Yoshioka R, Kiyosawa H et al (2000) X-Linked syndrome of polyendocrinopathy, immune dysfunction, and diarrhea maps to Xp11.23–Xq13.3. *Am J Hum Genet* 66:461–468
 80. Powell BR, Buist NR, Stenzel P (1982) An X-linked syndrome of diarrhea, polyendocrinopathy, and fatal infection in infancy. *J Pediatr* 100:731–737
 81. Ziegler SF (2006) FOXP3: of mice and men. *Annu Rev Immunol* 24:209–226
 82. Moraes-Vasconcelos D, Costa-Carvalho BT, Torgerson TR, Ochs HD (2008) Primary immune deficiency disorders presenting as autoimmune diseases: IPEX and APECED. *J Clin Immunol* 28(Suppl 1):S11–S19
 83. Gambineri E, Torgerson TR, Ochs HD (2003) Immune dysregulation, polyendocrinopathy, enteropathy, and X-linked inheritance (IPEX), a syndrome of systemic autoimmunity caused by mutations of FOXP3, a critical regulator of T cell homeostasis. *Curr Opin Rheumatol* 15:430–435
 84. Nieves DS, Phipps RP, Pollock SJ et al (2004) Dermatologic and immunologic findings in the immune dysregulation, polyendocrinopathy, enteropathy, X-linked syndrome. *Arch Dermatol* 140:466–472
 85. Torgerson TR, Ochs HD (2007) Immune dysregulation, polyendocrinopathy, enteropathy, X-linked: forkhead box protein 3 mutations and lack of regulatory T cells. *J Allergy Clin Immunol* 120:744–750
 86. Heltzer ML, Choi JK, Ochs HD et al (2007) A potential screening tool for IPEX syndrome. *Pediatr Dev Pathol* 10:98–105
 87. Bennett CL, Ochs HD (2001) IPEX is a unique X-linked syndrome characterized by immune dysfunction, polyendocrinopathy, enteropathy, and a variety of autoimmune phenomena. *Curr Opin Pediatr* 13:533–538
 88. Costa-Carvalho BT, de Moraes-Pinto MI, de Almeida LC et al (2008) A remarkable depletion of both naïve CD4+ and CD8+ with high proportion of memory T cells in an IPEX infant with a FOXP3 mutation in the forkhead domain. *Scand J Immunol* 68:85–91
 89. Watanabe N, Wang YH, Lee HK et al (2005) Hassall's corpuscles instruct dendritic cells to induce CD4+ CD25+ regulatory T cells in human thymus. *Nature* 436:1181–1185
 90. Korzenik JR (2007) Is Crohn's disease due to defective immunity? *Gut* 56:2–5
 91. Thrasher AJ, Keep NH, Wientjes F, Segal AW (1994) Chronic granulomatous disease. *Biochim Biophys Acta* 1227:1–24
 92. Segal AW, Abo A (1993) The biochemical basis of the NADPH oxidase of phagocytes. *Trends Biochem Sci* 18:43–47
 93. Segal AW (2005) How neutrophils kill microbes. *Annu Rev Immunol* 23:197–223
 94. Segal BH, Romani LR (2009) Invasive aspergillosis in chronic granulomatous disease. *Med Mycol* 47(Suppl 1):S282–S290
 95. Stasia MJ, Li XJ (2008) Genetics and immunopathology of chronic granulomatous disease. *Semin Immunopathol* 30:209–235

96. Werlin SL, Chusid MJ, Caya J, Oechler HW (1982) Colitis in chronic granulomatous disease. *Gastroenterology* 82:328–331
97. Marciano BE, Rosenzweig SD, Kleiner DE et al (2004) Gastrointestinal involvement in chronic granulomatous disease. *Pediatrics* 114:462–468
98. Westerberg LS, Meelu P, Baptista M et al (2010) Activating WASP mutations associated with X-linked neutropenia result in enhanced actin polymerization, altered cytoskeletal responses, and genomic instability in lymphocytes. *J Exp Med* 207:1145–1152
99. Thrasher AJ, Burns SO (2010) WASP: a key immunological multitasker. *Nat Rev Immunol* 10:182–192
100. Symons M, Derry JM, Karlak B et al (1996) Wiskott–Aldrich syndrome protein, a novel effector for the GTPase CDC42Hs, is implicated in actin polymerization. *Cell* 84:723–734
101. Blanchoin L, Amann KJ, Higgs HN et al (2000) Direct observation of dendritic actin filament networks nucleated by Arp2/3 complex and WASP/Scar proteins. *Nature* 404:1007–1011
102. Ochs HD, Thrasher AJ (2006) The Wiskott–Aldrich syndrome. *J Allergy Clin Immunol* 117:725–738
103. Villa A, Notarangelo L, Macchi P et al (1995) X-linked thrombocytopenia and Wiskott–Aldrich syndrome are allelic diseases with mutations in the WASP gene. *Nat Genet* 9:414–417
104. Notarangelo LD, Mazza C, Giliani S et al (2002) Missense mutations of the WASP gene cause intermittent X-linked thrombocytopenia. *Blood* 99:2268–2269
105. Devriendt K, Kim AS, Mathijs G et al (2001) Constitutively activating mutation in WASP causes X-linked severe congenital neutropenia. *Nat Genet* 27:313–317
106. Wiskott A (1937) Familiärer, angeborener Morbus Werlhofii? *Monatsschr Kinderheilkd* 68:212–216
107. Aldrich RA, Steinberg AG, Campbell DC (1954) Pedigree demonstrating a sex-linked recessive condition characterized by draining ears, eczematoid dermatitis and bloody diarrhea. *Pediatrics* 13:133–139
108. Sullivan KE, Mullen CA, Blaese RM, Winkelstein JA (1994) A multiinstitutional survey of the Wiskott–Aldrich syndrome. *J Pediatr* 125:876–885
109. Cannioto Z, Berti I, Martellosi S et al (2009) IBD and IBD mimicking enterocolitis in children younger than 2 years of age. *Eur J Pediatr* 168:149–155
110. Dupuis-Girod S, Medioni J, Haddad E et al (2003) Autoimmunity in Wiskott–Aldrich syndrome: risk factors, clinical features, and outcome in a single-center cohort of 55 patients. *Pediatrics* 111(5 Pt 1):e622–e627
111. Rohr J, Pannicke U, Döring M et al (2010) Chronic inflammatory bowel disease as key manifestation of atypical ARTEMIS deficiency. *J Clin Immunol* 30:314–320